

# Tubulin Tyrosine Ligase and Stathmin Compete for Tubulin Binding *In Vitro*

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## Abstract

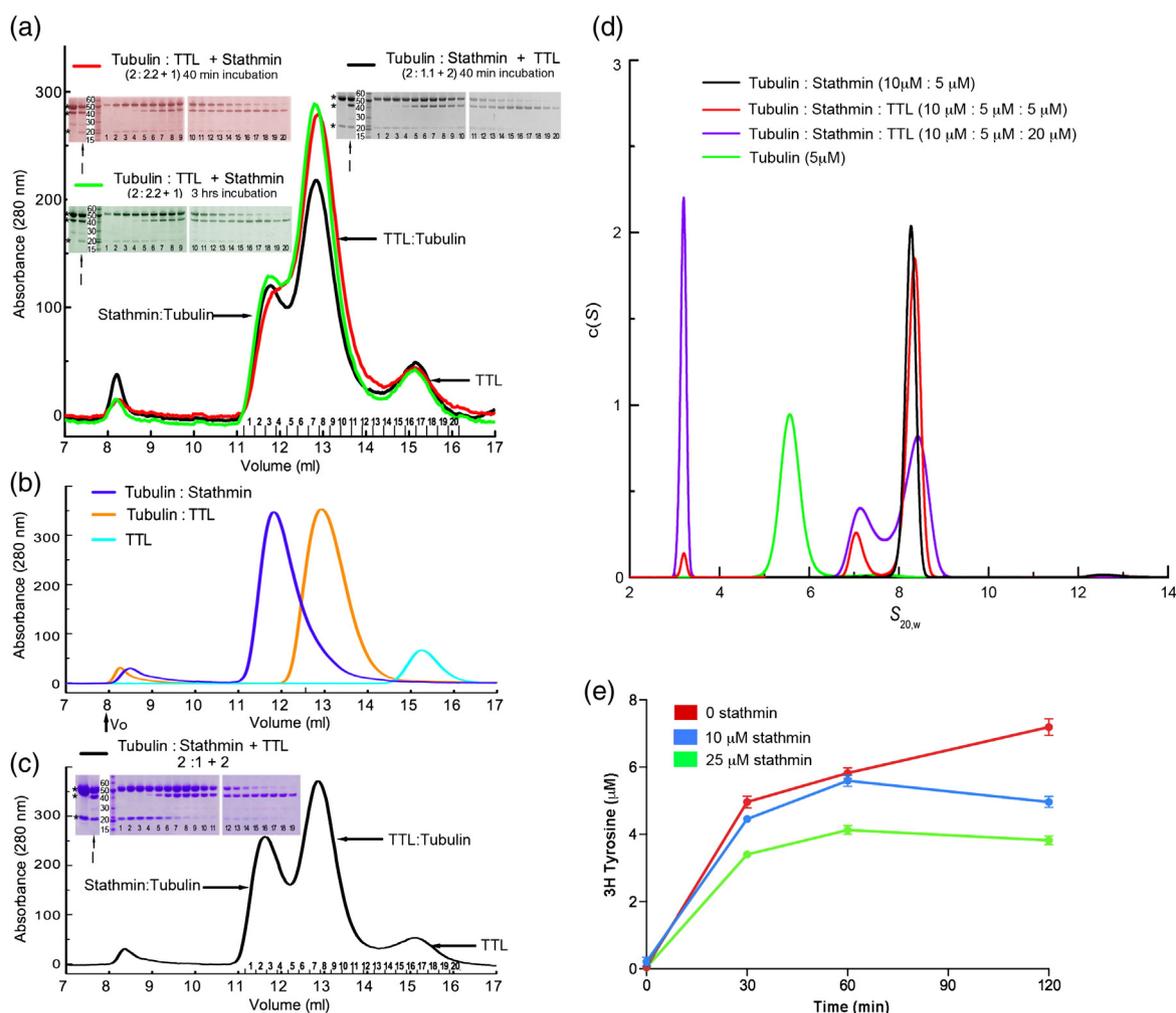
Tubulin partition between soluble and polymeric forms is tightly regulated in cells. Stathmin and tubulin tyrosine ligase (TTL) each form stable complexes with tubulin and inhibit tubulin polymerization. Here we explore the mutual relationship between these proteins *in vitro* and demonstrate that full-length stathmin and TTL compete for binding to tubulin and fail to make a stable tubulin:stathmin:TTL triple complex in solution. Moreover, stathmin depresses TTL tubulin tyrosination activity *in vitro*. These results suggest either that TTL and stathmin have a partially overlapping footprint on the tubulin dimer or that stathmin induces a tubulin conformation incompatible with stable TTL binding.

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The morphology and dynamics of the microtubule network are tightly regulated through interactions with a vast array of cellular factors as well as the post-translational modification of tubulin.<sup>1,2</sup> Stathmin/oncoprotein 18<sup>a</sup> is a highly conserved microtubule dynamics regulator that is ubiquitously expressed at high levels.<sup>3,4</sup> Tubulin tyrosine ligase (TTL) adds a C-terminal tyrosine to  $\alpha$ -tubulin. TTL is essential and also ubiquitously expressed. Both TTL and stathmin were isolated from cell extracts in complex with tubulin<sup>4-6</sup> and were each shown to interact preferentially with unpolymerized tubulin subunits,<sup>3,4,7</sup> however, with different stoichiometries. TTL forms a 1:1 complex with tubulin<sup>5,8</sup> while stathmin forms a complex by sequestering two tubulin dimers.<sup>3</sup> TTL and stathmin each inhibit tubulin growth rates *in vitro* and *in vivo*.<sup>3,4,8,9</sup> Solution studies revealed that TTL binds at the edge of the tubulin dimer, making contacts mostly with  $\alpha$ -tubulin and possibly the  $\alpha\beta$  protomer junction where it can interfere with both longitudinal and lateral tubulin polymerization interfaces.<sup>8</sup> Consistent with this, TTL inhibits tubulin polymerization *in vitro*.<sup>8</sup>

Stathmin strings two curved, longitudinally arranged tubulin dimers ( $\alpha 1 \beta 1-\alpha 2 \beta 2$ ) using an extended  $\alpha$ -helix that makes interactions with structural elements in the nucleotide binding (the S3-H3 loop, the C-terminus of helix H4 and the N-terminus of the H4-S5 turn) and C-terminal domains (the turn immediately preceding C-terminal helix H12 that leads into the disordered tubulin tails) of each of the  $\alpha$ - and  $\beta$ -tubulin protomers. In addition, stathmin caps the  $\alpha$ -tubulin longitudinal interface at one end of the complex *via* an N-terminal  $\beta$ -hairpin that extends a preexisting  $\beta$ -sheet in  $\alpha$ -tubulin containing conserved residues important for longitudinal interactions.<sup>10</sup> Stathmin is disordered in solution in the absence of tubulin.<sup>3</sup>

Since stathmin and TTL were each isolated complexed with tubulin and both inhibit tubulin polymerization, we investigated whether binding of full-length stathmin and TTL to tubulin are mutually exclusive and whether TTL can form a triple complex with tubulin:stathmin and tyrosinate it efficiently. Figure 1 shows that, under conditions where stable tubulin:stathmin and tubulin:TTL complexes are



**Fig. 1.** Solution studies of TTL and stathmin binding to tubulin. (a) Complex formation between TTL, stathmin and tubulin in various orders of addition of TTL and stathmin monitored by size-exclusion chromatography. Tubulin and TTL were pre-incubated for 40 min, followed by incubation with stathmin for another 40 min resulting in final concentrations of 47, 52 and 23  $\mu\text{M}$  for tubulin, TTL and stathmin, respectively (red), or 3 h (green); tubulin and stathmin were pre-incubated for 2 h followed by incubation with TTL for 40 min, resulting in final concentrations of 44, 24 and 44  $\mu\text{M}$  for tubulin, stathmin and TTL, respectively (black). The gels correspond to the indicated peak fractions. Molar ratios of the components are indicated. Asterisks (\*) indicate SDS-PAGE mobility of tubulin, TTL and stathmin (top to bottom); I is input. (b) Size-exclusion chromatograms of binary complexes of tubulin:stathmin (blue) injected at 2:1.1 molar ratio, tubulin:TTL (orange) at 1:1.1 molar ratio and TTL alone (cyan). (c) Size-exclusion chromatography of tubulin, stathmin and TTL mixed at indicated molar ratios; three times as much sample as in (a) was injected onto the same column. (d) AUC analyses of tubulin:stathmin:TTL mixtures showing the absence of a tubulin:stathmin:TTL triple complex and the competition between TTL and stathmin for tubulin binding. (e) Tubulin tyrosination by TTL is depressed by stathmin. We pre-incubated 10  $\mu\text{M}$  tubulin with 0, 10 and 25  $\mu\text{M}$  stathmin for 40 min before starting the tyrosination reaction (Supplementary Material).

detected by size-exclusion chromatography when all three components are incubated together, no tubulin:stathmin:TTL triple complex is detected while binary tubulin:stathmin: and tubulin:TTL complexes are (Fig. 1a and b). This result is independent of the order of addition of the three components. Pre-incubation of stathmin and tubulin for various times followed by addition of TTL or *vice versa* did not alter the result, implying that the formation of a stable

tubulin:stathmin:TTL triple complex in solution is unfavorable over the binary complexes, regardless of the order of addition of the components. The inability to detect a triple complex is unlikely due to dilution during chromatography because the same result holds when three times as much sample is loaded on the same column (Fig. 1c). The concentration of soluble tubulin in the cytoplasm is ~5–10  $\mu\text{M}$ .<sup>11</sup> Stathmin levels comparable to those of

tubulin have been detected (Refs. 4 and 9 and references therein) while TTL levels have been estimated to be lower in the cell<sup>5,6</sup> (~1:150 TTL:tubulin molar ratio<sup>5</sup>). Even at concentrations more than five times higher than these, we fail to observe any triple complex formation in our assay. We complemented our gel-filtration results with analytical ultracentrifugation (AUC) studies of tubulin:stathmin:TTL mixtures (Fig. 1d). The sedimentation coefficient distributions [ $c(s)$ ] of tubulin:stathmin complexes to which increasing amounts of TTL were added show three peaks, a small  $s$  value peak representing free TTL,<sup>8</sup> a peak with an  $s$  value of 7.1 representing tubulin:TTL and a large  $s$  value peak of 8.3 representing the tubulin:stathmin complex. No peaks with higher  $s$  values were detected, consistent with the absence of a tubulin:stathmin:TTL triple complex under these conditions. Addition of 5 and 20  $\mu$ M TTL is accompanied by a 7% and 48% decrease in the area of the stathmin:tubulin peak, respectively (Fig. 1d), indicating the competition between TTL and stathmin for tubulin binding. Consistent with our size-exclusion and AUC data that show a competition between stathmin and TTL for binding to tubulin, stathmin mildly inhibits tubulin tyrosination by TTL in a dose-dependent manner (Fig. 1e). Together, these results suggest either that stathmin and TTL compete for a partially overlapping binding site on tubulin or that stathmin induces a tubulin conformation that is not conducive to stable binding by TTL. While this manuscript was in review, a crystal structure of a TTL:tubulin:RB3 complex (the latter being a stathmin-like homolog) was reported.<sup>12</sup> It shows one copy of a TTL molecule binding to two copies of the tubulin dimer in the tubulin:RB3 complex despite the fact that the tubulin:RB3 complex appears to contain two equivalent TTL binding sites. However, the authors did not present data supporting the existence and stoichiometry of such a complex in solution (Supplementary Materials). The interplay between TTL and stathmin activities *in vivo* remains to be explored.

Supplementary data to this article can be found online at <http://dx.10.1016/j.jmb.2013.04.017>

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## Abbreviations used:

AUC, analytical ultracentrifugation;  
TTL, tubulin tyrosine ligase.

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